**Supplementary table 1.** Standard protocol for double sequential immunofluorescent staining of NET components in paraffin-embedded tissue.

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| **Fixation** | 1. Fix MSGB in 1% formaldehyde solution for 6 hours at room temperature.
2. Wash biopsies 3 times with distilled water for 10 min each.
3. Add 70% ethanol; biopsies can be stored at 4 °C until they are dehydrated.
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| **Dehydration** | 1. Dehydrate the biopsies using three grades of alcohol in the following sequence: once in 70% ethanol for 10 min, 3 times in 95% ethanol for 10 min each and 3 times in 100% ethanol for 10 min each.
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| **Clearing**  | 1. Clear biopsies 3 times with 100% xylene for 10 min each.
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| **paraffin embedding** | 1. Immerse biopsies into paraffin 3 times at 60 °C for 30 min each.
2. Put biopsies into embedding cassettes and fill the mold with paraffin to make the paraffin blocks.
3. Allow paraffin to solidify and remove embedding cassettes.
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| **Tissue sectioning** | 1. Place the tissue block in a microtome and cut multiple tissue sections of 4-µm in thickness with two intervals of 25 µm.
2. Let the tissue slices float on the surface of the water (water bath ready at 45 °C).
3. Use silanized glass slides to catch the floating tissue sections.
4. Incubate the tissue sections on slides at 37 °C overnight in order to bind the tissue to the glass. After that, glass slides can be stored at 4 °C until use.
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| **Rehydration** | 1. Place glass slides into a coupling jar and rinse them in the solutions used for dehydration and clearing in the following order: 3 times in 100% xylene for 5 min each, 2 times in 100% ethanol for 5 min each, once in 95% ethanol for 5 min, once in 70% ethanol for 5 min, and finally once in distilled water for 5 min.
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| **Antigen retrieval** | 1. Pre-heat food-steamer or water bath with a coupling jar containing heat-induced epitope retrieval buffer (sodium citrate pH: 6) until T° reaches 92 °C.
2. Submerse glass slides into coupling jar, put the lid on and incubate for 25 min.
3. Remove the coupling jar and leave it at room temperature and allow sections on the glass slides to sit for 15 min to cool.
4. Wash the glass slides once with distilled water for 5 min and once with PBS 1X (pH 7.4) for 5 min.
5. Dry the edges of the tissue sections with filter paper and draw a circle around the sections with a hydrophobic pen to create a barrier.
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| **Immunofluorescence stain**  | 1. Place the slides in a humidified chamber and do double immunostaining protocol with sequential incubations following the next steps:
2. Add blocking buffer (casein 0.25% in PBS 1X) and incubate at room temperature for 1 hour.
3. Prepare the 1st primary antibody in a proper dilution in PBS 1X: anti-neutrophil elastase diluted 1/25 (sc-55549).
4. Remove blocking buffer and incubate the tissue sections with the 1st primary antibody at 4°C overnight.
5. Wash tissue sections three times with PBS 1X for 10 min each.
6. Prepare the 1st secondary antibody in PBS 1X: goat anti-mouse IgG alexa fluor 488 diluted 1/200 (A-11001 Thermofisher).
7. Incubate tissue sections with the 1st secondary antibody at room temperature for 1 hour in the dark.
8. Wash tissue sections three times with PBS 1X for 10 min each.
9. Incubate tissue sections in blocking buffer at room temperature for 1 hour in the dark.
10. Remove blocking buffer and add the 2nd primary antibody diluted in PBS1X: anti-myeloperoxidase antibody 1/50 (ab9535) or anti-TRIM21/Ro/SS-A antibody 1/50 (ab91423) or anti-La/SS-B antibody 1/50 (ab124932).
11. Incubate tissue sections with the 2nd primary antibody at room temperature for 2 hours in the dark.
12. Wash tissue sections three times with PBS 1X for 10 min each.
13. Prepare the 2nd secondary antibody and hoechst (fluorescent dye used to stain nucleic acids) in PBS 1X: goat anti-rabbit IgG alexa fluor 546 diluted 1/200 (A-11010 Thermofisher) and hoechst 33342 diluted 1/250 (H3570 Thermofisher).
14. Incubate tissue sections with the 2nd secondary antibody and hoechst at room temperature for 1 hour in the dark.
15. Wash tissue sections three times with PBS 1X for 10 min each in the dark.
16. Wash once with deionized water for 5 min.
17. Add a drop of mounting medium (Entellan 107961 Sigma-Aldrich) to the tissue section and put a coverslip over it while avoiding bubble formation.
18. Store in the dark at 4°C.
19. Display the target using fluorescence microscopy.
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