Short communication

Shigatoxigenic and atypical enteropathogenic Escherichia coli in fish for human consumption

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ABSTRACT

Shigatoxigenic and enteropathogenic Escherichia coli with virulence and multidrug resistance profile were isolated from Nile tilapia. This study finding is of great importance to public health because they help understand this pathogen epidemiology in fish and demonstrate how these animals can transmit E. coli related diseases to humans.

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Escherichia coli (E. coli) is not a natural inhabitant of the fish microbiota, nevertheless, it can be isolated from these animals gut due to its presence in contaminated aquatic environments.1 It is worth noticing that this microorganism have pathogenic strains standing out as emerging zoonotic potential, as well as shigatoxigenic (STEC) and enteropathogenic (EPEC) E. coli. STEC strains produce the shiga toxin (Stx), which is its main virulence factor. There are two classes of shiga toxin, Stx1 and Stx2, with the last one presenting seven subtypes.2 The EPEC may be either typical or atypical, with the atypical strains do not carrying virulence factor that encodes the bundle-forming pilus (bfp), but it carries the eae gene, that is located at the locus of enterocyte effacement (LEE), which is a pathogenicity island, that promote attaching and effacing lesions (A/E). The ability to induce A/E lesions is mediated by genes located on the LEE, as well as additional ones that are outside of it.3

Several studies have analyzed STEC and EPEC, and their virulence in humans,4 cattle,5 sheep,6 pigs,7 and buffaloes.8 However, only a few studies have analyzed the presence of STEC and EPEC in fish9,10 and, of those, none has detected presence of adhesion and ESBL genes. In addition, none has performed the stx2 subtyping in STEC strains from fish. In this regard, this pioneer study aimed to compare the prevalence

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of STEC and EPEC strains in intensively farmed fish and free-living fish; as well as to detect their virulence and antibiotic resistant profile and analyze their genetic similarity looking for how these fishes contribute to humans infections.

The Ethics Committee on Animal Use (CEUA) approved this study under the protocol number 04076/14. Primers used are described in Table 1. The samples were collected from the fish species Oreochromis niloticus, from six different fish farms and three ranches located at northeast region of Sao Paulo state. A total of 472 samples were collected. Three hundred and seventy three (373) samples were obtained from fish farm animals and of these, 275 were from stools, 80 from muscles and 18 from the nurseries water. The other 99 samples were obtained from free-living fish, these 90 from stools and nine from the river water. Samples were transferred to tubes containing BHI broth (Brain Heart Infusion) and after an incubation period, the DNA were extracted by thermal lysis according to Borges.7

Screening for the detection of STEC and EPEC strain were based on, the stx1, stx2 and eae genes detection by multiplex PCR.7 When one of these genes were detected, individual colonies from each sample were tested by PCR to isolate STEC and EPEC strains according to the protocol available at www.apzec.ca/en/APZEC/Protocols/pdfs/ECL_PCR_Protocol.pdf. This methodology is in accordance to the OIE Reference Laboratory for Escherichia coli (Ecl – Faculté de Médecine Vétérinaire, Université de Montréal). Isolates were further submitted to another PCR to detect others virulence genes as follow: bfpA, eahX, saa, iha, toxB, efa1, lpfA0113, lpfA0157/OI-141, lpfA0157/OI-154, astA and paa genes. The Stx2 variants analysis was performed by stx2 subtyping according to Scheutz.7

The antimicrobial susceptibility test was performed using the disc diffusion method.30 The antimicrobials chosen were the ones most used in fish farming and which are important for the detection of resistance genes dissemination. In this regard, drugs tested were ampicillin (10 μg), cefalothin (30 μg), streptomycin (10 μg), gentamicin (10 μg), ciprofloxacin (5 μg), chloramphenicol (30 μg), tetracycline (30 μg), nitrofurantoin (300 μg), nalidixic acid (30 μg), sulfamethoxazole and trimethoprim (25 μg), ceftriaxone (30 μg), cefoxitin (30 μg), kanamycin (30 μg), norfloxacin (10 μg), enrofloxacin (5 μg) (Oxoid). In addition, E. coli isolates were screened for extended-spectrum beta-lactamase (ESBL) genes for the blaCTX-M genotype groups 1, 2, 8, 9 and 25, the blatTEM, and the blasHV.11

Phylogenetic E. coli groups’ classification was performed according to the methodology proposed by Clermont.12 Serotyping was performed at the E. coli Reference Center (ECRC) at Pennsylvania State University. The O somatic antigen were determine by agglutination plates, also the PCR-RFLP of fiC gene, which encodes flagella, were performed to determine the H flagella antigen. Somatic antigens used were O1 to O187, with the exception of O31, O47, O67, O72, O94, O122 and the flagellar antigens used were H1 to H49, except H17, since these serogroups still not have been designated.

The isolates were also characterized by PFGE pattern of the PulseNet protocol as described by Ribot.13 Briefly, the chromosomal DNA was digested with XbaI and the electrophoresis conditions were an initial time of 2.2 s and an end time of 54.2 ± s in a gradient of 6V and the gels were electrophoresed for 24 h. The fragment similarities were compared using the Dice coefficient and the dendrogram was constructed by neighbor-joining grouping using BioNumerics software (Applied Maths, Sint-Martens-Latem, Belgium). MLST was performed following the Achtman’s scheme (http://mlst.ucc.ie/mlst/dbs/Ecoli), through the sequencing of the PCR amplification products of the adk, fumC, gyrB, icd, mdh, purA, and recA genes. The generated sequences were trimmed and analyzed by the Phred/Phrap/Consed software package.

All the results are shown in Fig. 1. Of the 373 analyzed samples from the fish farm, one (0.2%), from stools, tested positive for a STEC related gene (isolate 125F5). Of the 99 free-living fish analyzed samples, six (6%), also from stools, were positive for at least one of the STEC or EPEC related genes (isolates 6F8, 9F8, 10F8, 12F8, 24F8 and 30F8). In addition, all six isolates were collected from the same location, and the stx1, stx2 and eae genes were detected. None of the muscle or water samples tested were positive for the STEC or EPEC markers investigated. Isolates from the fish farms were positive for eahX, lpfA0113 and saa virulence genes. Also, strains from the free-living fish presented astA, eahX, lpfA0113, saa, efa1 and paa genes. Regarding Stx2 toxin variants, the subtypes stx2a, stx2c and stx2d were observed at the same isolate.

Fig. 1 – A dendrogram representing the genetic similarity relationship and virulence indicators in STEC and aEPEC isolates from fish.

<table>
<thead>
<tr>
<th>Isolate</th>
<th>stx</th>
<th>eae</th>
<th>lpfA0113</th>
<th>eahX</th>
<th>saa</th>
<th>efa1</th>
<th>paa</th>
<th>astA</th>
<th>Phylotypea</th>
<th>Serotype†</th>
<th>stx2</th>
<th>ST‡</th>
<th>CC§</th>
<th>Resistance</th>
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<td>9F8</td>
<td>1</td>
<td>2</td>
<td>+</td>
<td></td>
<td></td>
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<td></td>
<td>B1</td>
<td>O39:H14</td>
<td>301</td>
<td>165</td>
<td>AMP, CFL, TET</td>
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<td>+</td>
<td>+</td>
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<td></td>
<td></td>
<td>B1</td>
<td>ONF:HNT</td>
<td>230</td>
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<td></td>
<td></td>
<td>B1</td>
<td>ONF:H18</td>
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<td></td>
<td>B1</td>
<td>O55:HNT</td>
<td>a,d</td>
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<td></td>
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<td></td>
<td>B1</td>
<td>O116:HNT</td>
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<td>29</td>
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<td>B1</td>
<td>O116:H36</td>
<td>58</td>
<td>155</td>
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* F5: fecal sample from fish farm animal; F8: fecal sample from wild animal; 10F8 is the only aEPEC strain and the others, STEC strains.
† NT: non-typeable
‡ ST: Sequence type from MLST
§ CC: Complement from MLST ST
§§ SUT-sulfamethoxazole and trimethoprim.
**Table 1 – Information about the primers used in PCR reactions.**

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Table 1 – (Continued)

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^a [http://mlst.warwick.ac.uk/mlst/dbs/Ecoli/documents/primerColi.html](http://mlst.warwick.ac.uk/mlst/dbs/Ecoli/documents/primerColi.html)

From the 15 antimicrobial drugs tested, the isolates originated from the fish farm animals were resistant to 14 of them, while the isolates from the free-living fish were resistant to three antimicrobials and no ESBL genes were found. All of the STEC strains belong to group B1 and the aEPEC strain to group A as in accord with the classification of Clermont.22 From seven isolates analyzed by serotyping, three were nontypeable for the O antigen, and three isolates were nontypeable for the H antigen. Thus, the groups detected were O55, O39, O116, H14, H18 and H36; and their serotype is shown in Fig. 1. Seven isolates possessed a heterogeneous profile, by PFGE analysis, and seven distinct sequence types (STs) with four clonal groups (CCs) detected by the MLST technique.

Although STEC and aEPEC strains isolated from fish are not natural inhabitants of its microbiota, these strains can colonize the fish through a contaminated environment of which they live.1 In both establishments that these positive strains were isolated, presence of cattle were observed around the nurseries and rivers. It is important to notice that bovine is considered the main reservoir of pathogenic E. coli.26

Moreover, in this study, all muscle and water samples were negative for the presence of STEC or EPEC. This result should not be taken lightly, because the samples were collected by dissecting the animal using aseptic conditions so that muscles samples were carefully separated from the intestinal content, which it does not occur at the fishermen or slaughterhouses daily practice. Commonly, a cut is made between the anus and the fish’ head, releasing all of its intestinal contents and, in the process, contaminating the muscle. In this regard, according to Kim,21 pathogenic E. coli can enter the human food chain mainly through food contamination. Moreover, none of the water sample tested positive for STEC or EPEC and this was associated with the large water flow at the nurseries and rivers.

These pathogens have already detected in Brazil4,6,8,14, and in other countries such as United States,15 Argentina,16 Netherlands,26 Iran,18 Tunisia,19 and Australia.20 These studies, as well as the present one, are fundamental to understand these pathogens epidemiology, since they have great importance in animal and public health.

In this study, one STEC strain with eae gene also contained several other genes, efa1, ehxA, lpfA0113 and paa, which have been associated with cases of diarrhea.21,22 The presence of saa gene was shown to be closely related to the presence of the ehxA gene in STEC strains devoid of eae gene, regardless of their serotype.4,23 In an STEC isolated, the presence of astA gene, which is important for pathogenesis of diarrhea and plays a key role in this strain’ virulence,24 was observed. Also, the lpfA0113 gene was shown in previous reports to have a high prevalence in STEC isolated from different animals.4 Regarding Stx2 toxin variants, the stx2a, stx2c and stx2d subtypes were observed in the same isolate; this combination is very unusual and makes this strain fairly unique and with aggravated virulence. The presence of stx2a, stx2c and stx2d subtype are often associated with the hemorrhagic colitis and with hemolytic uremic syndrome.2

The quinolones, tetracyclines, aminoglycosides and amphenicols are the most commonly used antimicrobials in fish farming25 and thus, it may explain the multiresistance observed in the fish farm isolates. All of the tested quinolones, tetracyclines, aminoglycosides and amphenicols were ineffective. Although it was observed multiresistance, no ESBL genes were found. The ESBL would confer an even greater risk for the raw fish consuming population, because these bacteria’s can produce an enzyme that are able to hydrolyze the beta lactam ring of penicillins, cephalosporins and aztreonam, thus conferring resistance to these antimicrobials.23 However, this multiresistance profile shows a phenotypic response from the isolates, and due to the selective pressure originated from the abusive use of broad-spectrum cephalosporins, according to data obtained in the present study, ESBL strains can emerge rapidly, as it is already observed in other animal productions.

According to the phylogenetic results of this study, it is confirmed that tropical populations can harbor strains of group A and B1 preferentially, which may be one of the factors that could explain tropical countries higher diarrhea frequency.27 Although much is reported about the O157 strains, non-O157 STEC strains are the most prevalent in animals and food. For this reason, chances of humans becoming infected by these strains are large, indicating the importance of this serogroups to public health.5 Therefore, the O116 serogroup observed in the present study is relevant due to the fact that it is often associated with severe human disease. Furthermore, the strain with the flagellar antigen H18 which contains the stx2 but not the eae gene, has also shown frequent association with infections in animals.28 Moreover, the ONT: H18 serotype in eae negative and saa positive strains, similarly to the ones in this study, has previously been detected in pathogenic E. coli in cattle,29 thus emphasizing the fact that these animals were, likely, the source of this pathogen infection in fish.

The genetic diversity analysis showed that, although the isolates belonged to the same bacterial species, they had genetic diversities that were highlighted though the PFGE technique. The same was observed with the MSLT data, indicating that although they had a common ancestral origin, the transference of genetic information, though time, made this
isolates very diverse, thus explaining their distinct phylogenet- 
cic classification. 
Ours results shows that fish can harbor an important com-
bination of Stx2 subtypes and putative adhesions genes. Also, 
it draws attention to the fact that the indiscriminate use of 
antibiotics in fish farming has the potential to endan-
ger consumer health through the dissemination of antibiotic 
resistance genes. And finally, it highlights the role of STEC and 
aEPEC as foodborne pathogens in fish for human consump-

Acknowledgments

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References

Escherichia coli in fresh water fish, Jenynsia multidentata and 
sequence-based protocol for subtyping shiga toxins and stan-
enteropathogenic Escherichia coli infection and prolonged 
Distribution of virulence profiles related to new toxins and 
putative adhesins in shiga toxin – producing Escherichia coli 
isolated from diverse sources in Brazil. FEMS Microbiol Lett. 
5. Brusa V, Restovich V, Galli L, et al. Isolation and 
characterization of non-0157 Shiga toxin-producing 
Escherichia coli from beef carcasses, cuts and trimmings of 
pathogenic Escherichia coli in healthy, pasture-raised sheep 
on farms and at the abattoir in Brazil. Vet Microbiol. 
2013;169:89–95.
7. Borges CA, Beraldo LG, Maluta RP, et al. Shiga toxigenic and 
atypical enteropathogenic Escherichia coli in the feces and 
carcasses of slaughtered pigs. Foodborne Pathog Dis. 
toxigenic (STEC) and enteropathogenic (EPEC) Escherichia coli 
9. Manna SK, Das R, Manna C. Microbiological quality of 
finfish and shellfish with special reference to shiga toxin-producing 
10. Ribeiro LF, Barbosa MMC, Rezende Pinto F, et al. Shiga 
toxigenic and enteropathogenic Escherichia coli in water and 
fish from pay-to-fish ponds. Lett Appl Microbiol. 
set of multiplex PCR assays for the detection of genes encoding 
important β-lactamases in Enterobacteriaceae. J Antimicrob 
Escherichia coli phylo-typing method revisited: improvement of 
specificity and detection of new phylo-groups. Environ 
13. Ribot EM, Fair MA, Gautom R, et al. Standardization of 
pulsed-field gel electrophoresis protocols for the subtyping of 
Escherichia coli O157:H7, Salmonella, and Shigella for 
toxin-producing Escherichia coli in drinking water supplies of 
north Paraná State, Brazil. J Appl Microbiol. 
2013;114:1230–1239.
15. Herman KM, Hall AJ, Gould LH. Outbreaks attributed to fresh 
2015;143:3011–3021.
characterization of Escherichia coli virulence genes isolated 
from diseased pigs in the central region of Argentina. Vet 
17. Paton AW, Paton JC. Multiplex PCR for direct detection of 
Shiga toxigenic Escherichia coli strains producing the novel 
Escherichia coli pathotypes in north and north-west provinces of 
resistance and molecular characterization of virulence 
genes, phylogenetic groups of Escherichia coli isolated from 
diarrheic and healthy camel-calves in Tunisia. Comp Immunol 
Shiga toxin-producing Escherichia coli infection in Australia. 
virulence genes linked with diarrhea due to atypical 
Enteropathogenic Escherichia coli by DNA microarray analysis 
non-locus of enterocyte effacement pathogenic 
island-related genes in Escherichia coli carrying eae from 
patients with diarrhea and healthy indi-viduals in Japan. J 
Escherichia coli heat-stable enterotoxin 1 represents another 
subfamily of E. coli heat-stable toxin. Proc Natl Acad Sci U S A. 
1993;90:3093–3097.
association of Escherichia coli AIDA-USTb, rather than EAST1 
pathotype, with diarrhea in piglets and antibiotic sensitivity 
25. Rigos G, Troisi GM. Antibacterial agents in Mediterranean 
fish farming: a synopsis of drug pharmacokinetics in 
important euryhaline fish species and possible 
environmental implications. Rev Fish Biol Fish. 2005;15: 
53–55.
of Human Infections with Shiga Toxin-Producing Escherichia coli 
(STEC) to Livestock Sources and Identification of Source-Specific 
Health; 2017.
27. Escobar-Páramo P, Grenet K, Menac’h AL, et al. Large-scale 
population structure of human commensal Escherichia coli 
28. Eklund M, Scheutz F, Siitonen A. Clinical isolates of 
non-O157 shiga toxin-producing Escherichia coli: serotypes, 
virulence characteristics, and molecular profiles of strains of 
virulence genes, and intimin types of Shiga toxin-producing 
Escherichia coli (STEC) and enteropathogenic E. coli (EPEC) 
isolated from calves in São Paulo, Brazil. Int J Food Microbiol. 


